RESEARCH



Detection of *Trypanosoma evansi* in jaguars (*Panthera onca*): insights from the Brazilian Pantanal wetland

Renata Fagundes-Moreira¹ · Vinicius Baggio-Souza¹ · Joares Adenilson May-Junior^{1,2,3} · Laura Berger¹ · Lina Crespo Bilhalva¹ · Adeyldes Oliveira Reis¹ · Leonardo Sartorello² · Lilian E. Rampim² · Marcos Antônio Bezerra-Santos⁴ · Domenico Otranto^{4,5} · João Fabio Soares¹

Received: 21 November 2023 / Accepted: 16 December 2023 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2024

Abstract

Trypanosoma evansi is a widespread and neglected zoonotic parasite that affects domestic and wild animals, causing a disease commonly known as "surra." The Brazilian Pantanal wetland is recognized as an enzootic area for this protozoan, yet recognizing the importance of reservoir hosts also in order to prevent zoonotic outbreaks. This study aimed to assess the occurrence of *T. evansi* in jaguars (*Panthera onca*) from the Brazilian Pantanal wetland and explore associated clinical and hematological manifestations. A total of 42 animals were screened by PCR and sequenced for species identification when positive. *Trypanosoma evansi* was detected in six free-ranging jaguars (six positive animals of 42 captures and 16 recaptures), representing the first molecular evidence of such infection in this animal species. Our findings suggest that jaguars may act as reservoir hosts of *T. evansi* in the Brazilian Pantanal wetland. The better understanding of the role of wildlife in the epidemiology of *T. evansi* is also of importance to future reintroduction and translocation programs toward wildlife conservation efforts.

Keywords Hemoparasite · Protozoan · Wild felids · Tripanossomiasis · Brazil

Introduction

Trypanosoma evansi, the etiological agent of a neglected disease known as "surra" or in Portuguese "mal das cadeiras," affects a wide range of domestic and wild animals (Franke et al. 1994; Silva et al. 1995; Herrera et al. 2004), with sporadic cases also reported in humans (Joshi et al., 2005; Powar et al. 2006). Previous studies have documented

Handling Editor: Una Ryan

João Fabio Soares joao.soares@ufrgs.br

- ¹ Laboratório de Protozoologia e Rickettsioses Vetoriais, Faculty of Veterinary, Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, Rio Grande do Sul, Brazil
- ² Onçafari Association, São Paulo, Brazil
- ³ Panthera Corporation, New York, USA
- ⁴ Department of Veterinary Medicine, University of Bari, Valenzano, Italy
- ⁵ Department of Veterinary Clinical Sciences, City University of Hong Kong, Kowloon Tong, Hong Kong

that many animal species, such as cattle (Franke et al. 1994), horses (Rodrigues et al. 2005), dogs (Echeverria et al. 2019; Nguyen et al., 2021), tigers (Upadhye and Dhoot, 2000), camels (Desquesnes et al. 2008), and bats (Herrera et al. 2004), and even humans may be susceptible to *T. evansi* infection. In addition, a zoo outbreak was reported in India, which many felid species, including jaguars (*Panthera onca*), died due to *T. evansi* (Sinha et al., 1971).

The Brazilian Pantanal biome has been identified as an enzootic area for *T. evansi* (Nunes et al. 1993). Therefore, understanding the ecology of this protozoan in the region is advocated, especially considering the potential wild reservoir hosts (Herrera et al. 2004).

The ecosystem complexity and its biodiversity require a comprehensive approach to disease ecology, including studies about potential disease reservoirs among both common and endangered wildlife species. The jaguar, as one of the apex predators in South America, merits particular attention, and although its susceptibility to the infection has been demonstrated in captive animals (Sinha et al., 1971; Khan et al., 2023), the role of free ranging jaguars in the transmission dynamics of *T. evansi* remain understudied. Under the above

circumstances, this study aimed to assess the occurrence of *T. evansi* infection in free-ranging jaguars within the Brazilian Pantanal wetland.

Material and methods

The study was conducted in the municipality of Miranda, Mato Grosso do Sul state, Brazil, within the Pantanal biome, between 2013 and 2023 (Fig. 1). Jaguars were captured using foot snares (May-Junior et al. 2021) to collect biological samples and set GPS/VHF radio collars. The radio collars had drop-off system after 12 months, so recaptures were not necessary to remove it. In all animals, we assessed the health parameters, including mucosal color, capillary refill, hematocrit, blood smear, rectal temperature, and weight (kg); also, radio or GPS collars were set when possible, according to both collar availability and jaguar size. Only a young female jaguar was not set a radio collar, as she was young and too small (50 kg). Ethical procedures were approved by the Ministry of Environment under license number #42093-1.

Blood samples were collected and processed using the PureLink® Genomic DNA Mini Kit (InvitrogenTM, Carlsbad, CA, USA) to extract genomic DNA (200 μ L volume) following the manufacturer's instructions. Molecular detection of *T. evansi* involved the use of two specific primer protocols. Firstly, an assay was caried out to amplify a 315-bp

fragment following Ventura et al. (2002). Subsequently, an assay was conducted to amplify a ~ 540-bp ITS-1 fragment, as described by Desquesnes et al. (2001). Blood samples from experimentally infected mice and naturally infected dogs were used as positive controls. In order to evaluate the quantity and quality of the extracted DNA, a NanoDropTM spectrophotometer at an absorbance of 260/280 nm was used.

One random sample was selected to retrieve sequence. Amplicons of the expected size were purified and sequenced in both directions using the Big Dye Terminator v.3.1 chemistry in a 3130 Genetic Analyzer (Applied Biosystems, California, USA) equipped with an automated sequencer (ABI-PRISM 377). The resulting sequence was aligned using the fast Fourier transform algorithm in MAFFT (Katoh et al. 2019) and subsequently compared with reference sequences available in the GenBank database using the Basic Local Alignment Search Tool (BLAST). The sequence was submitted to the GenBank database under the accession number: OR797837.

Results

Six (four females and two males) out of 58 captures (42 animals and 16 recaptures) scored positive at PCR (Table 1) (Supplementary Table 1), representing an occurrence of 14.28% (6/42). At the BLAST analysis, the

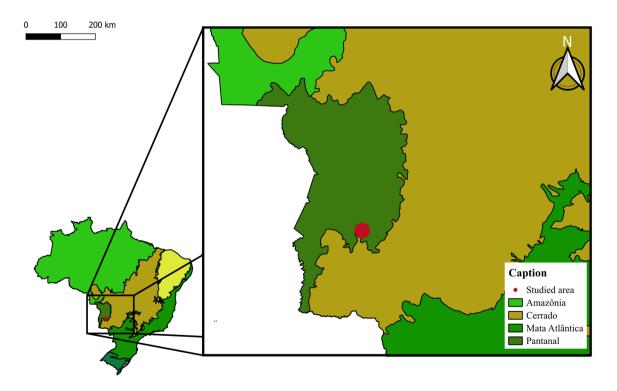


Fig. 1 Geographical location of the studied area in the Brazilian territory

Table 1 Captures and recaptures of Trypanosoma evansi positive free-ranging jaguars (Panthera onca)

Sample	Gender	Capture		Weight (Kg)	PCV (%)	Detection of
		Month	Year			T. evansi
bPon424	Female	September	2017	92	nt	+
bPon440	Female	June	2014	110	38	_
		January	2017	100	nt	_
		November	2018	86.4	38	+
bPon495	Male	July	2017	~ 120	nt	+
		June	2020	127	nt	_
bPon498	Female	September	2017	50.7	nt	+
bPon509	Female	November	2018	60	42	+
bPon522	Male	February	2021	96.2	40	+

Kg, kilogram; PCV, packed cell volume; %, percent; nt, not tested

evaluated sequence showed 100% nucleotide identity with T. evansi (Accession number: AB551922.1, AY912279.1, AF306775.1). The jaguars did not suffer for major clinical changes at the time of capture. However, two females (bPon 424 and bPon 440) had capillary refill in 2 (bPon 424) and 1.5 s (bPon 440) and presented a senile aspect with white hair and loss of muscle mass in their hind limbs, probably associated with aging, as bPon 424 was more than 10 years old and bPon 440, more than 12 years old. Jaguars maintained a regular PCV (Table 1), according to the parameters for the species in the studied region (40.7 \pm 5.0—calculus made from 37 monitored jaguars-data not shown). All blood smears evaluated through optical microscopy were negative for Trypanosoma sp.

Discussion

In this study, we report the occurrence of T. evansi in freeranging jaguars from the Pantanal wetland in Brazil. While the role of this animal species in the epidemiology of this parasite is unknown, the finding herein reported is of importance since their involvement in the maintenance of T. evansi in nature.

The absence of clinical signs compatible with T. evansi infection in animals herein evaluated contrasts with findings from captive jaguars (Sinha et al. 1971), and domestic cats, that shows clinical signs and positive in blood smears (Priyowidodo et al. 2023), suggesting that free-ranging individuals may exhibit a distinct response to this protozoan infection. Immunotolerance to trypanosomes in carnivores remains a complex and understudied phenomenon. However, extrinsic factors, such as the exposure to high challenges through predation of infected prey and invertebrate vectors, have been associated with resistance (Murray et al. 1982; Kasozi et al. 2021). In the context of our study population of free-ranging jaguars, it is likely that they exhibit trypanotolerance, mainly considering Pantanal region is an enzootic area for T. evansi (Herrera et al. 2004). In a similar scenario, Serengeti lions, which are frequently exposed to multiple trypanosome species through tsetse fly bites and the consumption of infected meat, have demonstrated cross-immunity to Trypanosoma brucei, enabling them to eliminate the infection after exposure (Welburn et al. 2008). Furthermore, it is worth noting that one male (bPon 495), initially tested positive upon capture, scored negative during a 3-year interval recapture. This observation suggests either the suppression of the infection or a reduction of trypanosomiasis to undetectable levels by PCR. This highlights the complexities of pathogen-host interactions and underscores the need for thorough species-specific investigations. In addition, the jaguar population of this study presents a high prevalence of infection by Cytauxzoon sp. (Fagundes-Moreira et al. 2022). The synergism between infections of distinct pathogens, associated with environmental changes, had important effects on the populations of African large felids (Munson et al. 2008).

One of the possible transmission routes to jaguars may be the ingestion of reservoir preys, once oral transmission was already suggested in the Indian zoo outbreak (Sinha et al. 1971, Khan et al., 2023) and proved in dogs and mice (Raina et al. 1985; Bazolli et al. 2002); additionally, leopards (Panthera pardus) are reservoirs to T. brucei and were also suspected to be orally infected by its prey (Anderson et al. 2011). For example, tapirs (Tapirus terrestris) and capybaras (*Hydrochaerus hydrochaeris*) are common prey for jaguars, being also considered reservoirs of T. evansi in the Pantanal wetlands, with high parasitemia recorded in capybaras (Franke et al. 1994; Herrera et al. 2004; Rademaker et al. 2009; Fundación Rewilding Argentina, 2020). The transmission of T. evansi to domestic animals is commonly associated with tabanid bites (Desquesnes et al. 2005, Kamidi et al., 2017), but it may also be the case for wildlife reservoirs. For example, previous studies

demonstrated that tabanid species (e.g., *Tabanus importunus*, *Tabanus occidentalis*), associated with *T. evansi*, were more prevalent during the rainy season, from September to the first days of March (Silva et al., 1995; Barros, 2001), which also coincides with capybaras breeding period (Aldana-Domínguez et al. 2002). Accordingly, in this study, five out of six *T. evansi* positive jaguars were diagnosed during the rainy season. Therefore, the increase of vectors and the availability of "easier" prey (younger and susceptible capybaras) acting as reservoirs may represent an important factor in *T. evansi* transmission to jaguars.

The implications of these findings are far-reaching for wildlife conservation efforts, especially considering the critical role of jaguars in the ecological balance of the Pantanal wetland ecosystem. Importantly, our results should be incorporated into risk assessments for reintroduction or translocation programs of jaguars and other South American wildlife to prevent accidental pathogen introduction to a susceptible population and potential disease outbreaks (Viggers et al. 1993).

Conclusion

Our findings from the Pantanal demonstrate the presence of subclinical *T. evansi* in South American free-ranging jaguars. As our understanding of the relationships between pathogens, hosts, and the environment continues to grow, the importance of interdisciplinary research in developing effective conservation strategies remains undeniable. Therefore, further studies on the impact of *T. evansi* on jaguars are advocated to better understand the risks associated and guide the safeguarding of biodiversity and long-term survival of this endangered species.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00436-023-08101-0.

Acknowledgements The authors would like to thank the Refúgio Ecológico Caiman, Onçafari Association, Log Nature, Zoetis Inc., and Leatherman for the support in the conduction of the study. In addition, the authors thank Viviane Noll Louzada Flores and Livia Perles (University of Bari, Italy) for the support on the molecular analysis.

Author contributions RFM: Conceptualization, Investigation, Methodology, Writing—original draft preparation. VBS: Formal Analysis, Methodology, Writing—review and editing. JAMJ: Investigation, Methodology, Writing—review and editing. LB: Conceptualization, Formal Analysis, Investigation, Methodology. LCB: Investigation, Methodology. AOR: Investigation, Methodology. LS: Investigation, Methodology. LER: Investigation, Methodology. MABS: Investigation, Methodology, Writing—review and editing. DO: Conceptualization, Methodology, Supervision, Writing— review and editing. JFS: Conceptualization, Investigation, Methodology, Project administration, Supervision, Writing—review and editing. Funding This work received support from Fundação de Amparo à Pesquisa do Rio Grande do Sul (FAPERGS) (Finance code:19/2551–0001842–8), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) (Finance code 001). The coauthors MABS and DO were supported by the EU funding within the MUR PNRR Extended Partnership initiative on Emerging Infectious Diseases (Project no. PE00000007, INF-ACT). The coauthor JFS is funded by CNPq (Grant #312576/2021-8). This article is based on the development of activities carried out by RFM during the Programa Institucional de Internacionalização (CAPES-PRINT) sandwich doctoral period at the Department of Veterinary Medicine, University of Bari, Italy.

Data availability No datasets were generated or analyzed during the current study.

Declarations

Ethics approval The procedures herein described were conducted in accordance with the Brazilian Institute of the Environment and Renewable Natural Resources-IBAMA (Authorization n. 42093-1) and the Research Committee of the Federal University of Rio Grande do Sul-Compesq (Authorization n. 38198).

Consent to participate Not applicable.

Consent for publication Not applicable.

Competing interests The authors declare no competing interests.

References

- Aldana-Domínguez J, Forero-M J, Betancur J, Cavelier J (2002) Dinámica y estructura de la población de chiguiros (*Hydrochaeris hydrochaeris*: rodentia hydrochaeridae) de Caño Limón, Arauca, Colombia. Zoología 24(2):445–458
- Anderson NE, Mubanga J, Fevre EM, Picozzi K, Eisler MC, Thomas R, Welburn SC (2011) Characterisation of the Wildlife Reservoir Community for Human and Animal Trypanosomiasis in the Luangwa Valley, Zambia. PLoS Negl Trop Dis 5(6):e1211. https:// doi.org/10.1371/journal.pntd.0001211
- Barros ATM (2001) Seasonality and relative abundance of Tabanidae (Diptera) captured on horses in the Pantanal, Brazil. Mem Inst Oswaldo Cruz 96:917–923
- Bazolli RS, Marques LC, Machado RZ, Aquino LPC, Alessi AC, Camacho AA (2002) Oral transmission of *Trypanosoma evansi* in dogs. Ars Vet 18(2):148–152
- Desquesnes M (2005) Livestock trypanosomoses and their vectors in Latin America. Trans R Soc Trop Med Hyg. https://academic.oup. com/trstmh/article/99/9/716/1886005
- Desquesnes M, McLaughlin G, Zoungrana A, Dávila AM (2001) Detection and identification of *Trypanosoma* of African livestock through a single PCR based on internal transcribed spacer 1 of rDNA. Int J Parasitol 31:610–614
- Desquesnes M, Bossard G, Patrel D, Herder S, Patout O, Lepetitcolin E, Thevenon S, Berthier D, Pavlovic D, Brugidou R, Jacquiet P, Schelcher F, Faye B, Touratier L, Cuny G (2008) First outbreak of *Trypanosoma evansi* in camels in metropolitan France. Vet Rec 162(23):750–752. https://doi.org/10.1136/vr.162.23.750
- Echeverria JT, Soares RL, Crepaldi BA et al (2019) Clinical and therapeutic aspects of an outbreak of canine trypanosomiasis. Rev Bras Parasitol Vet 28:320–324. https://doi.org/10.1590/S1984-29612 019018

- Fagundes-Moreira R, Souza UA, May-Junior JA, Baggio-Souza V, Berger L, Wagner PGC, Mazim FD, Peters FB, Favarini MO, Tortato MA, Albano APN, Fagundes DD, Haberfeld MB, Sartorelo LR, Ranpim LE, Fragoso CE, Girotto-Soares A, Martins TF, Valle SF, Soares JF (2022) Epidemiological compatibility of *Amblyomma sculptum* as possible vector and *Panthera onca* as reservoir of *Cytauxzoon* spp. in Midwestern Brazil. Ticks Tick borne Dis 13(6):102021. https://doi.org/10.1016/j.ttbdis.2022.102021
- Franke CR, Greiner M, Mehlitz D (1994) Investigations on naturally occurring *Trypanosoma evansi* infections in horses, cattle, dogs and capybaras (*Hydrochaeris hydrochaeris*) in Pantanal de Poconé (Mato Grosso, Brazil). Acta Trop 58(2):159–169. https://doi.org/ 10.1016/0001-706X(94)90055-8
- Fundación Rewilding Argentina (2020). Tapires y el "Mal de las caderas" en Parque Iberá, https://www.rewildingargentina.org/tapir es_mal_caderas_ibera/
- Herrera HM, Rademaker V, Abreu UGP (2004) Enzootiology of Trypanosoma evansi in Pantanal, Brazil. Vet Parasitol 125:263–275
- Joshi PP, Shegokar VR, Powar RM, Herder S (2005) Human trypanosomiasis caused by *Trypanosoma evansi* in India: the first case report. Am J Trop Med Hyg 73:491–495
- Kamidi CM, Saarman NP, Dion K, Mireji PO, Ouma C, Murilla G, Aksoy S, Schnaufer A, Caccone A (2017) Multiple evolutionary origins of *Trypanosoma evansi* in Kenya. PLoS Negl Trop Dis 11(9):e0005895. https://doi.org/10.1371/journal.pntd.0005895
- Kasozi KI, Zirintunda G, Ssempijja F, Buyinza B, Alzahrani KJ et al (2021) Epidemiology of trypanosomiasis in wildlife—implications for humans at the wildlife interface in Africa. Front Vet Sci 8:1–15. https://doi.org/10.3389/fvets.2021.621699
- Katoh K, Rozewicki J, Yamada KD (2019) MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. Briefings in Bioinformatics 20(4):1160–1166. https:// doi.org/10.1093/bib/bbx108
- Khan MA, Shabir S, Azeem S, Gill W, Ashraf K, Azhar M, Rashid I, Ashraf M, Avais M, Ahmad AS, Younas M, Badshah A, Ahmad S, Akbar H (2023) Documentation of *Trypanosoma evansi* in captive tigers and lions in Punjab (2016-2018), Pakistan. J Zoo Wildl Med 53(4):823–831. https://doi.org/10.1638/2021-0053
- May-Junior JA, Fagundes-Moreira R, Baggio-Souza V, De Almeida, BA, Haberfeld MB, Sartorelo LR, Ranpim LE, Fragoso CE, Soares JF (2021) Dermatobiosis in *Panthera onca*: First description and multinomial logistic regression to estimate and predict parasitism in captured wild animals. Rev Bras Parasitol Vet 30(1). https://doi.org/10.1590/s1984-29612021003
- Munson L, Terio KA, Kock R, Mlengeya T, Roelke ME, Dubovi E, Summers B, Sinclair ARE, Packer C (2008) Climate extremes promote fatal co-infections during canine distemper epidemics in African Lions. PLOS ONE 3(6):e2545. https://doi.org/10.1371/ journal.pone.0002545
- Murray M, Morrison WI, Whitelaw DD (1982) Host susceptibility to African trypanosomiasis: trypanotolerance. Adv Parasitol 21:1– 68. https://doi.org/10.1016/S0065-308X(08)60274-2
- Nguyen VL, Iatta R, Manoj RRS, Colella V, Bezerra-Santos MA, Mendoza-Roldan JA, Otranto D (2021) Molecular detection of *Trypanosoma evansi* in dogs from India and Southeast Asia. Act Trop 220:105935

- Nunes VLB, Oshiro ET, Dorval MEC, Garcia LAM, Da Silva AAP, Bogliolo AR (1993) Investigação Epidemiológica Sobre *Trypanosoma* (Trypanozoon) *evansi* no pantanal sul-mato-grossense -Estudo de reservatórios. Rev Bras Parasitol Vet 2(1):41–44
- Powar RM, Shegokar VR, Joshi PP, Dani VS, Tankhiwale NS et al (2006) A rare case of human trypanosomiasis caused by Trypanosoma evansi. Indian J Med Microbiol 24(1):72
- Priyowidodo D, Sahara A, Prastowo J, Nurcahyo W, Firdausy LW (2023) Detection of *Trypanosoma evansi* in a naturally infected cat in Indonesia using bioassay and molecular techniques. Vet World 16(4):828
- Rademaker V, Herrera HM, Raffel TR, D'Andrea PS, Freitas TPT et al (2009) What is the role of small rodents in the transmission cycle of *Trypanosoma cruzi* and *Trypanosoma evansi* (Kinetoplastida Trypanosomatidae)? A study case in the Brazilian Pantanal. Acta Trop 111(2):102–107. https://doi.org/10.1016/j.actatropica.2009. 02.006
- Raina AK, Kumar R, Sridhar VSR, Singh RP (1985) Oral transmission of *Trypanosoma evansi* infection in dogs and mice. Vet Parasitol 18(1):67–69
- Rodrigues A, Costa MM, de Macedo GC (2005) Surtos de tripanossomíase por *Trypanosoma evansi* em equinos no Rio Grande do Sul: aspectos epidemiológicos, clínicos, hematológicos e patológicos. Pesq Vet Bras 25(4):239–249
- Silva RAMS, Barros ATM, Herrera HM (1995) Foyers Trypanosomiens Dus à *Trypanosoma evansi* Dans Le Pantanal, Brésil. Une Approche Préliminaire Sur Les Facteurs de Risque. Rev élev méd vét pays trop 48(4):315–319. https://doi.org/10.19182/remvt.9432
- Sinha PK, Mukherjee GS, Das MS, Lahiri RK (1971) Outbreak of Trypanosomiasis evansi amongst tigers and jaguars in the zoological garden, Calcutta. Indian Vet J 48(3):306–310
- Upadhye SV, Dhoot VM (2000) Trypanosomiasis in a tiger (*Panthera tigris*). Zoos' Print Journal 15(8)
- Ventura RM, Takeda GF, Silva RA, Nunes VL, Buck GA, Teixeira MM (2002) Genetic relatedness among *Trypanosoma evansi* stocks by random amplification of polymorphic DNA and evaluation of a synapomorphic DNA fragment for species-specific diagnosis. Int J Parasitol 32:53–63
- Viggers KL, Lindenmayer DB, Spratt DM (1993) The importance of disease in reintroduction programmes. Wildl Res 20(5):687–698
- Welburn S, Picozzi K, Coleman PG, Packer C (2008) Patterns in ageseroprevalence consistent with acquired immunity against *Trypa*nosoma brucei in Serengeti lions. PLoS Negl Trop Dis 2(12):1–6. https://doi.org/10.1371/journal.pntd.0000347

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.